

# WRAP UP

# Annual Meeting – ISEV2017 Toronto, Canada 18-21 May 2017 www.isev.org

# **NEWS IN**



#### FLOW CYTOMETRY

- hrFC
- hollow silica beads
- nanoFCM
- NP-40 detergent



#### **EV ISOLATION**



- EV sorting

- ExoDisk

- Microfluidics



**EV CHARACTERIZATION** 

- live imaging
- pHluorin
- EV uptake
- Different MVBs
- Single vesicle analysis

#### **FUNCTIONAL STUDIES**

-EV uptake

-"cloud of vesicles"

-EVs and Cardiovascular system

-EVs and Pericardial fluid

-EVs and AMI



#### **EV BIOPHYSICS AND BIOCHEMISTRY**



EV diameter ↑, then stochastic events ↑ EV molecular imaging – wireless network EV labeling by location: - inside - transmembrane outer membrane inner membrane

EV detergent sensitivity may vary between subpopulations?

Interference of optiprep use with lowry based protein assay

Molecular heterogeneity

Circulating plasma older and newer EVs?

Selection mechanism(s)

Different MVBs  $\rightarrow$  different exosomes

ARRDC1 -recruitment of MVB to PM

CD9+ EVs are independent of RABs

EV internalization is inhibited by latrunculin

Nsmase2 control secretion of miR via exosomes

Live cell imaging of exosomes: https://pdfs.semanticscholar.org/9128/842ec2e6351feafc4ad5633ddcafe4b39756.pdf

Fusion with PM through vSNARE/ tSNARE - how are EVs are escaping EE?

**EV UPTAKE** 



#### ENDOCYTOSIS

- A. Clathrin dependent
- B. Caveolin dependent
- C. Lipid raft mediated (FLOT may be involved)

Stripping proteins from EVs reduces their uptake (eg. Proteinase K treatment)

#### Proteins involved in uptake:

 → tetraspannins: CD81, CD9, CD63
→ integrins and immunoglobulins: CD51, CD61
→ proteoglycans: HSPGs (Christianson 2013)
→ lectins Abs again DCSIGN reduces EV uptake-DEC205 PHAGOCYTOSIS

Through PS and other lipids



Inhibition of binding: tripsinidase, neutralizing Abs

Inhibition of endocytosis: Dynasore, Pitstop + list of drugs

Zomer et al, Nat Protocols, 2016; Mulcahy et al, 2014 JEV

#### Lipids in exosomes



hr proteomic and lipidomic analysis Haraszti et al 2016 JEV

DRARPAD.com

#### Single vesicle analysis



Proposed isolation method: microfluidics

**Confocal microscopy**: mitochondria DSRED detects vesicles containing mitochondrias

EV quant vs NTA: 0.99 correlation, can detect small vesicles up to 50 nm down; detect multiple surface markers on EVs





#### **EV CHARACTERIZATION**



- EV quantification: EV/ Cell/ Hour
- Saturation: reach concentration  $\rightarrow$  dilution : concentration dependent curve (±co-culture experiments)
- Chris Gardiner : NTA/TRPS/DLS do not distinguish the protein aggregates from EVs
- EV best measured in isotonic aqueous medium
- To prevent loss of particles over time in diluted suspensions: dilute immediately before use
- Reduce sample error: 5-20 short measurements are better instead of 1 long
- Check regularly the instrument calibration!
- DLS can be useful when combined with SEC
- TEM: wide EM + 1-2 close view ± immunogold labeling; CryoSEM Koffman ez al, 2017
- DOT BLOT permeabilization: Tween20, inside vs outside of EVS: Sung et al, 2017 Cell Adh Migr
- Surface biothinylation followed by MS

#### **EV ISOLATION**



**C. Thery**: no pressure based filtration isolation: could break large EVs and create artificial small EVs tagging proteins may alter the effects of EVs

Membrane filtration	Acoustic separation	Immuno-magnetic separation
-low purity -it can process large volumes -combing filters→ reduces yield	-no mechanical or chemical stress on EVs -SAW (surface acustic wave) Lee et al, ACS NANO 2015	-bead quoted with Ab → highly specific ! selection of Abs; mixture of Abs (CD9, CD63, CD81)
Affinity separation	ODG	-low yield Jeoung et al, ACS NANO 2016
-up to 12 marker in one sample -Ab based	EV + HDL together	ExoDISK
-ELISA and nPLEX	Remnant OPTIPREP can be successfully removed by SEC	lab-on-a-disc with 2 nanofilters 20-600 nm range
<b>qEV EXOSOME ISOLA</b>		Wo et al, ACS Nano 2017

Sucrose gradient density centrifugation: EVs may shrink +  $\uparrow$  density was measured of the EV fractions  $\rightarrow$  use Optiprep instead

#### FLOW CYTOMETRY (1) evflowcytometry.org



High frequency of small signals are buried in noise

Hollow silica beads: RI:1,46; do not sediment, membrane thickness: 5 nm

MIEV Flow Cyt Checklist

nanoFCM: detection 1-10 epitope vs. Astrios nanoFACS detection 30-200 epitopes (vs. conventional FACSCalibur +200 epitope)

Plasma EV epitope profile with hrFC biolegend +300Ab

Synthetic MP: Smith et al Circulation 2017

EVs from cardiomyocytes can be sorted and isolated (GFP+EVs) Das et al

non fluorescent particles affect fluorescent labeled EVs: High concentration of unstained particles negatively impact the fluorescent thresholding

NanoFACS: sorted EVs are biologically viable and can be used for RNAseq

Zabeo et al 2017 JEV

Kormelink et al, 2016; Stomer et al, 2016

#### hr FLOW CYTOMETRY (2)



Apogee A50+

Astrios-EQ nanoFACS

#### - 80 nm Fluorescent particle detection



- High speed detection

#### - EV sorting



Jones et al 2016

nanoFCM – prototype May 2017

- Highly dedicated setup
- Limited dynamic range
- Best detection limit



www.nanofcm.com

#### EV RNA (1)



exosome-rna.com

2 different exRNA signatures released by a single cell type

There is not one miRNA, that is constantly secreted in exosome

Y RNAs are present in EVs and they are altered in DC activation

Treating EVs with UV for 220 sec: successfully decreases RNA without damaging proteins



Cambier et al, 2017 Lässer et al, 2017





How does bovine serum-derived RNA affect EV-RNA analysis?

Observation of miRNA species in EV pellets of cell-conditioned medium that could not be detected in parent cell Eg. Liver hsa-miR-122 in glioblastoma cell supernatant

- 1. Depletion of (EV) RNA from bovine serum prior to cell culture may not be efficient
- 2. Bovine RNAs can map to murine or human genome (up to 10% alignment for both miR, mRNA, snoRNA)

#### **APOPTOTIC BODIES**



IAV induces apoptosis and apoptotic bodies production

Epithelial cell line A549 treated with AB derived from IAV infected THP-1 cells induced infection in vitro

THP-1 cell derived AB could induce the same infection in vivo in mice

AB from infected cell can elicit inflm response and IAV specific T cell response

Can we impair viral infection by influencing the AB production?

Inhibiting AB disassembly with drug  $x \rightarrow$  partial restorement of cell disassembly

#### EVs and the CARDIOVASCULAR SYSTEM



tRNAs are increased in primary myocardiocytes derived EVs

Diabetes promotes release of miR503 enriched MP from endothelium

CD34+ exosomes: are pro-angiogenic and have an increased has-miR-130a, 126a and 92a In hypoxia CD34+ exosome conc is increased

CD34+ exosomes induce angiogenesis in vivo (mice) loss of miR-126a reduced the angiogenic potential of Cd34+ exosomes. Uptake of CD34+ exosomes injected in the heart: by endothelial (via SIRPA) and cardiomyocyte cells, and no uptake by fibroblast cells. These exosomes may induce the cell cycle in endothelial cells.

hsa-miR-21 activates cardiac fibroblasts + antagomir miR-21 as general anti-fibrotic drugs (Phase II clinical trial)

CVD alter neuronal targets of circulating EVs

CD151 and mucin 16 is increased with age in EVs

#### **EVs and ACUTE MYOCARDIAL INFARCTION**



After AMI 2-3 days: inflammatory monocytes M1 After AMI 4 days : M2 type monocytes

个VCAM1+ EVs Anti-VCAM1 reduces the EC migration (Boyden chamber in vitro) 个个TNF+ EVs

AMI modulates EV miRNA: 个 miR:126-3p, 126-5p, 26b-5p, 1472, 23a-3p, 151a-3p, 374b-5p

EC-EVs mobilize the monocytes from spleen in vivo mice

In human: Monocytes from spleen are mobilized after AMI

EVs reduce oxidative stress under hyperglycemic conditions



#### **EVs and the PERICARDIAL FLUID**



CABG diabetes pericardial fluid EVs (CABG non diabetes, MVR)

Exosomes: CD63, Alix, TSG101 + Beltrami et al, 2017

RNA:

→ Proangiogenic response: let-7b-5p; hsa-mir-21-5p; has-mir126-3p (relative to cel-miR-39)

- ightarrow increase apoptosis in EC
- → comparing microRNA signature with the miRNA pattern in piece of atrium and piece of aortic tissues MS:
- $\rightarrow \uparrow$  expression of apoptotic proteins in CABG DM PF
- $\rightarrow$  APAF1 protein (under control by let-7b-5p; protein presence validated by WB)

WB

CD63 APAF1 double positive EVs

Costanza Emanueli: The pericardial fluid exosomes as new cell-to-cell communicators worsening ischaemic heart disease in diabetes Andrew I.U. Shearn: Methodological considerations for nanoparticle tracking analysis (NTA) of neat biofluids obtained from cardiac surgery Sezin Aday: Bio-inspired synthetic exosomes carrying microRNA let-7b for post- ischemic vascular regeneration



Host- virus interaction in ocean

Algal bloom in ocean $\rightarrow$	50% of O2 production
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Emiliania huxleyi - are killed by EhV viruses (dsDNA, 500 genes)

Viral free lysate =VFL induces vesicle production, in these vesicles RNA and lipid profile identified, no DNA

In virus infected E. huxleyi derived EVs: different lipidomic profile: TAG and SL different small RNA profile: with protein targeting chromatin remodeling

EV: 1. provirus; 2. prohost

BODIPY stain for vesicles

EVs lead to increased stability of the virus  $\rightarrow$  EV/EhV ratio is  $\uparrow \rightarrow$  EhV half life is  $\uparrow$ 

Vesiclomics in the ocean Project: collection of EVs from the ocean in marine environment

Schatz et al, 2017 Nat Microbiology; Ziv et al, 2014; Rosenwarver et al, 2014



- Injection of EVs, interspecies
- In vitro and in vivo (animal models)
- One patient studied in GVH
- Experiments started for large scale production of therapeutic EVs
- Toxicity/immunogeneity/ safety
- MSC



#### **Keynote Speakers**



https://www.selectbiosciences.com/conferences/index.aspx?conf=EVE2017



# TOP 5: RBC-derived EVs role in cardiac remodeling in vivo

AMI: Mo/MF, Fibr, Endoth, RBC  $\rightarrow$  infiltration into heart  $\rightarrow$  heart remodeling  $\rightarrow$  Dysfunction in Kidney, Brain and Hematopoetic System

Cre-lox system: ROSA26: kidney receives cre from the heart EVs

Cardiomyocytes EVs in plasma are 5-8% RBC releases EVs with complement activation: possible mechanism to target EVs

EpoR-cre-mTmG mice have GFP+ RBC derived EVs

EVs from EPO-Cre RBC can transfer functional cre

RBC EVs communicate with heart (cardiomyocytes), kidney (prox tubular cells), spleen (splenic cells), lung

+ control experiment to exclude ectoptic expression (FC assay)



# **TOP 4: Circulating cancer** associated EVs

Α

Whole blood

В







Erythrocytes (~5×10e9/mL blood)

Leukocytes (~7×10e<sup>6</sup>/mL blood)

Thrombocytes (~3×10e8/mL blood)

Normal exosomes (~10e<sup>11</sup>/mL blood)

Tumor stroma exosomes (unknown)

Tumor exosomes (~0-5×10e<sup>10</sup>/mL blood\*)

Normal cfDNA (~5×10e9/mL blood)

Tumor cfDNA (~5×10e9/mL blood)

Ago2 associated miRNA (~5×10e9/mL blood)

Brock et al, 2015 HDL associated miRNA (~5×10e9/mL blood)



# **TOP 3: Imagestream<sup>x</sup> hrFC**





## **TOP 2: Imagestream<sup>x</sup> Mark II**





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K562 cell line, DAPI (purple), the lysosomes were stained with CD107a AF647 (red), and the endosomes were stained with CD71 FITC (green)



### **TOP 1: In vivo tracking of**

#### exosomes

Frederik Verweij, Institut Curie, Paris

Zebrafish placenta: yolk specific expression of CD63pHluorin

Active uptake of EVs by MF: initially are not degraded/ acidified

Extravasation of EVs towards interstitial fluid: form a major repository for exosomes



# Thank you for your attention!

# Extracellular vesicles or vesicles?

